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Design and Characterization of Floating Microspheres of Rabeprazole Sodium for Prolonged Gastric Retention

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ABSTRACT

Floating drug delivery system is one of the novel drug delivery system. Floating drug delivery system have a bulk density less than gastric fluids and so remain buoyant in the stomach without affecting gastric emptying rate for a prolonged period of time. The aim of the present study was to develop floating microspheres of Rabeprazole sodium(RPS), which belong to class of proton pump inhibitor. Floating microspheres of Rabeprazole were prepared by emulsion solvent evaporation method using HPMC K15M and ethyl cellulose as polymer. Six different formulations were developed. The floating microsphere was evaluated for angle of repose, particle size, percentage yield, in vitro buoyancy, incorporation efficiency, drug polymer compatibility (IR study), scanning electron microscopy, drug release and DSC(Differential Scanning calorimetry), X-Ray Diffraction(XRD) of microsphere. Results show that as the concentration of polymer increases it affects the particle size, percentage yield, in vitro buoyancy and drug release of microsphere. Formulations prepared with HPMC K15M exhibited excellent Micromeritic properties, percentage yield, in vitro buoyancy, incorporation efficiency and percentage drug release when compared to ethyl cellulose polymer. Results of our present study suggest that floating microsphere of Rabeprazole sodium can be successfully designed to develop controlled drug delivery which can reduce dosing frequency thus this formulation can be considered as an alternative to conventional dosage forms.

Key words: floating drug delivery systems, Rabeprazole sodium (RPS), incorporation efficiency, dosing frequency, DSC(Differential Scanning calorimetry) X-Ray Diffraction (XRD)

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INTRODUCTION

The oral route is considered as the most promising route of drug delivery. Conventional drug delivery system achieves as well as maintains the drug concentration within the therapeutically effective range needed for treatment, only when taken several times a day. This results in a significant fluctuation in drug levels. Recently, several technical advancements have led to the development of several novel drug delivery systems (NDDS) that could revolutionize method of medication and provide a number of therapeutic benefits. The most important objectives of these new drug delivery systems are: First, it would be single dose, which releases the active ingredient over an extended period of time. Second, it should deliver the active entity directly to the site of action, thus, minimizing or eliminating side effects¹. To overcome the limitations of conventional drug delivery system, floating drug delivery systems have been developed. Drugs that have narrow absorption window in the gastrointestinal tract (GIT) will have poor absorption. For these drugs, gastroretentive drug delivery systems offer the advantages in prolonging the gastric emptying time.

Floating drug delivery systems are among the several approaches that have been developed in order to increase the gastric residence time of the dosage forms^{2,3,4}. The floating drug delivery systems have been extensively used to improve therapy with several drugs. However during development process several difficulties are faced such as inability to restrain and localize the system within desired region of the GIT and its variable as per gastric emptying process. The variability may cause unpredictable bioavailability and time to achieve peak plasma levels. On the other hand, incorporation of the drug in a controlled release gastroretentive dosage forms (CR-GRDF) which can remain in the gastric region for several hr would significantly prolong the gastric residence time of drugs and improve bioavailability, reduce drug waste, and enhance the solubility of drugs that are less soluble in high pH environment⁵. This FDDS provides local delivery of drug in the form of microspheres to specific region like stomach and also shows better bioavailability and improved therapeutic activity and substantial benefits to patients.

Floating microspheres are gastro- retentive drug delivery systems based on non- effervescent approach. Hollow microspheres are in strict sense, spherical empty particles without core. These microspheres are characteristically free flowing powders consisting of proteins or synthetic polymers, ideally having a size range less than 200 micrometer. Gastroretentive floating microspheres are low density systems that have sufficient buoyancy to float over a gastric content and remain buoyant for prolonged period of time. The drug is released slowly at a

desired rate resulting in increased gastric retention with reduced fluctuations in plasma drug concentration. Floating microspheres to improve patient compliance by decreasing dosing frequency, better therapeutic effect of short half –life drugs can be achieved ⁶.

The present study involves formulation and in-vitro evaluation of floating microspheres of Rabepazole Sodium under gastro retentive drug delivery system for improving bioavailability by prolonging gastric retention time. In the present work Rabepazole sodium was chosen as the drug to be incorporated into the individual polymer like HPMC K15 M and EC. Rabepazole sodium is a drug prescribed to treat disorders caused by excess stomach acid. Available as the brand AcipHex. Rabepazole is a proton pump inhibitor similar to Prevacid and Prilosec. PPIs block an enzyme in the stomach from producing acid, significantly reducing gastric acid levels and allowing acid-related disorders to heal, as well as alleviating symptoms of chronic conditions drug that has been widely used in treating gastric and duodenal ulceration and also in Zollinger Ellison syndrome and reflux esophagitis ⁷. It is poorly absorbed from the lower GIT and has a short elimination half-life (2h). The Rabepazole has an oral bioavailability of approximately 52% ⁸. Rabepazole is less susceptible to the influence of genetic polymorphisms for CYP2C19, resulting in minor influence on its pharmacokinetics and pharmacodynamics ^{9,10}.

MATERIALS AND METHODS

Rabepazole sodium was obtained from Reddies lab Hyderabad. India as gift sample, HPMC K15 was provided by Shreeji chemicals, Mumbai. Ethyl cellulose was purchased from Rolex chemicals, Mumbai. Ethanol was obtained from S D fine chemical Ltd, Mumbai, Span 80, dichloromethane also obtained from S D fine chemical Ltd, Mumbai. All other ingredients were of analytical grade.

Preformulation studies

Selection of vehicle

The solubility of Rabepazole sodium was checked in various solvents like water and methanol, ethanol, chloroform and ethyl acetate, ether and n-hexane. Studies revealed that Rabepazole sodium was Rabepazole sodium was found to be very soluble in water and methanol, freely soluble in ethanol, chloroform and ethyl acetate and insoluble in ether and n-hexane⁷. The solubility was confirmed by analyzing the sample by quantitative determination by UV spectroscopy. Wavelength scan was done from 400-200 nm and maximum absorbance was found at 275.43nm.

Melting point determination

Melting point of RPS was determined by Open capillary method. The melting point of RPS was found to be 140-141°C.

Determination of λ_{\max}

A solution of RPS containing the concentration 10 µg/ ml was prepared in 0.1 M HCL and UV spectrum was taken using Shimadzu (UV-1800) double beam spectrophotometer. The solution was scanned in the range of 200 – 400 nm. Wavelength of maximum absorption of Rabeprazole Sodium in 0.1M HCL is found to be 275.43nm.

Drug polymer interaction (FTIR) study ¹¹

FTIR spectroscopy was performed on Fourier transformed infrared spectrophotometer (IR-Affinity-1, Shimadzu, Japan). The pellets of drug and potassium bromide were prepared by compressing the powders at 20 psi for 10 min on KBr-press and the spectra were scanned in the wave number range of 4000- 600 cm⁻¹.

Formulation studies

Preparation of Rabeprazole sodium floating microspheres ^{12, 13, 14, 15.}

Method used: Emulsification – solvent evaporation method Floating microspheres were prepared by solvent evaporation technique accurately weighed drug and individual polymer like HPMC K15 M and EC were dissolved in ethanol and dichloromethane (1:1) to form a homogenous polymer solution. This solution is poured in 100 ml light liquid paraffin containing 0.01% span 80 maintained at 30-40°C subsequently stirred at ranging agitation speed for 30 min to allow the volatile liquid to evaporate. The microspheres formed were filtered, washed with petroleum ether and dried in vacuum. The microspheres were then stored in a desiccator over fused calcium chloride.

Formulation design

Formulation design for Rabeprazole Sodium floating microspheres using different ratios of drug and polymers. (1, 2&3 HPMC K15M & 4, 5&6 Ethyl Cellulose) shown in Table1.

Sl. No	Formulation code	Average size (µm)
1	RPS1	73.32
2	RPS2	74.41
3	RPS3	78.52
4	RPS4	86.26
5	RPS5	90.83
6	RPS6	93.57

Evaluation of Rabeprazole Sodium Floating Microspheres

Drug polymer interaction (FTIR) study

FTIR spectroscopy was performed on Fourier transformed infrared spectrophotometer (IR-Affinity-1, Shimadzu, Japan). the spectra were scanned in the wave number range of 4000- 600 cm^{-1} . FTIR study was carried on RPS loaded microspheres.

Micromeritic properties

❖ Angle of repose

Angle of repose of different formulations was measured according to fixed funnel standing method ($n = 3$) $\theta = \tan^{-1}h / r$ where θ is the angle of repose, r is the radius, and h is the height

Scanning electron microscopy (S.E.M)

Scanning electron microscopy has been used to determine particle size distribution, surface topography, texture, and to examine the morphology of fractured or sectioned surface. SEM is probably the most commonly used method for characterizing drug delivery systems, owing in large to simplicity of sample preparation and ease of operation. SEM studies were carried out by using JEOL JSM T-330A scanning microscope (Japan). Dry RPS floating microspheres were placed on an electron microscope brass stub and coated with in an ion sputter. Picture of RPS floating microspheres were taken by random scanning of the stub.

Particle size analysis

Determination of average particle size of RPS floating microspheres was carried out by optical microscopy in which stage micrometer was employed. A minute quantity of RPS floating microspheres was spread on a clean glass slide and average size of 300 RPS floating microspheres was determined in each batch.

Percentage yield

The percentage yield of prepared RPS floating microspheres was determined by using the formula.

$$\text{Percentage yield} = \frac{\text{Practical yield}}{\text{Theoretical yield}} \times 100$$

Buoyancy percentage

Fifty milligrams of the floating microspheres were placed in 0.1M HCL, 100 ml containing 0.02 w/v% span 80. The mixture was stirred at 100 rpm in a magnetic stirrer. After 12 hrs, the layer of buoyant microspheres was pipette and separated by filtration. Particles in the sinking particulate layer were separated by filtration. Particles of both types were dried in a desiccators until constant weight.

$$\text{Buoyancy(\%)} = \frac{W_f}{W_f + W_s} \times 100$$

Where W_f and W_s are the weights of the floating and settled microspheres, respectively.

Determination of percentage drug entrapment (PDE)^{16,17}

Efficiency of drug entrapment for each batch was calculated in terms of percentage drug entrapment as per the following formula.

$$\text{PDE} = \frac{\text{Practical drug loading}}{\text{theoretical drug loading}} \times 100$$

➤ **Preparation of standard calibration curve of Rabeprazole sodium in 0.1M HCL**

- Scanning of Rabeprazole sodium by UV-spectrophotometer in 0.1M HCL

I Stock: 100 mg of RPS was accurately weighted into 100 ml volumetric flask, dissolved in 0.1M HCL and volume was made up with 0.1M HCL. II Stock: Pipette 1ml of above solution into another 10 ml volumetric flask and the volume was made with 0.1M HCL.

- **Procedure for calibration of Rabeprazole sodium in 0.1M HCL at λ_{max} 275.43nm**

From the RPS standard stock solution (1000 $\mu\text{g/ml}$), 1ml solution was diluted to 10 ml using 0.1M HCL solution to get concentrations of 100 $\mu\text{g/ml}$. from this solution, aliquots of, 0.5 ml, 1.0 ml, 1.5 ml, 2.0 ml, 2.5 ml, 3.0 ml from standard drug solution were diluted to 10 ml with 0.1M. The absorbance of these solutions was measured at 275.43nm 0.1M HCL as a blank.

❖ **Theoretical drug content**

Theoretical drug content was determined by calculation assuming that the entire RPS present in the polymer solution used gets entrapped in RPS floating microspheres, and no loss occurs at any stage of preparation of RPS floating microspheres

❖ **Practical drug content**

➤ **Procedure:** Practical drug content was analyzed by using the following procedure, weighed amount of RPS floating microspheres equivalent to eight mg of RPS floating microspheres was dissolved in 100 ml of 0.1 M HCL. This solution was kept overnight for the complete dissolution of the RPS floating microsphere in 0.1M HCL. This solution was filtered and further diluted to make a concentration of 10 $\mu\text{g/ml}$ solution. The absorbance of the solutions was measured at 275.43nm using double beam UV-Visible spectrophotometer against 0.1M HCL solution as blank and calculated for the percentage of drug present in the sample.

In vitro dissolution studies.

❖ **Procedure for Invitro dissolution study**

The release rate of RPS floating microspheres was determined by employing USP XXIII apparatus by rotating basket method. The dissolution test was performed using 900 ml 0.1M HCL, in $37 \pm 0.5^\circ\text{C}$ at 50 rpm. RPS floating microspheres equivalent to 20 mg were placed in a

basket to avoid floating of microspheres. A sample (5 ml) of the solution was withdrawn from the dissolution apparatus hourly for 12 hrs, and the samples were replaced with fresh dissolution medium. The samples were passed through Whatman filter paper and the absorbance of these solutions was measured at 275.43nm. Dissolution profiles of the formulations were analyzed by plotting drug release versus time plot. Data obtained was also subjected to kinetic treatment to understand release mechanism.

Kinetics of drug release

To examine the drug release kinetics and mechanism, the cumulative release data were fitted to models representing zero order (Q v/s t), first order [$\text{Log}(Q_0-Q)$ v/s t], Higuchi's square root of time (Q v/s $t^{1/2}$) and Korsmeyer Peppas double log plot ($\text{log } Q$ v/s $\text{log } t$) respectively, where Q is the cumulative percentage of drug released at time t and (Q_0-Q) is the cumulative percentage of drug remaining after time t .

In short, the results obtained from *in vitro* release studies were plotted in four kinetics models of data treatment as follows.

- Cumulative percentage drug release Vs. Time (zero order rate kinetics)
- Log cumulative percentage drug retained Vs. Time (first order rate kinetics)
- Cumulative percentage drug release Vs. \sqrt{T} (Higuchi's classical diffusion equation)
- Log of cumulative percentage drug release Vs. $\text{log } \text{Time}$ (Peppas exponential equation)

Differential Scanning Calorimetry (DSC)

The physical state of drug in the RPS floating microspheres was analyzed by DSC. The thermograms of RPS, RPS floating microspheres with different polymers were obtained at a scanning rate of 10°C/min conducted over a temperature range of 25–350°C, respectively.

X-Ray power Diffractometry (XRD) study¹⁸

X-ray diffractometry of the RPS and RPS microspheres were performed by a diffractometer using model (Joel JDX-8030, Japan) equipped with a graphite crystal monochromator (Cu-K α) radiations to observe the physical state of RPS in the microspheres

RESULTS AND DISCUSSION

Drug polymer interaction (FTIR) study

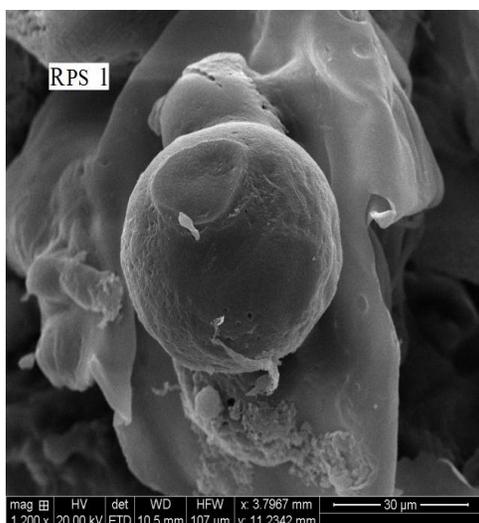
From the spectra of RPS, physical mixture of RPS and individual polymer, RPS loaded microspheres it was observed that all characteristic peaks of RPS were present in the combination spectrum, thus indicating compatibility of the RPS and polymer.

Angle of repose

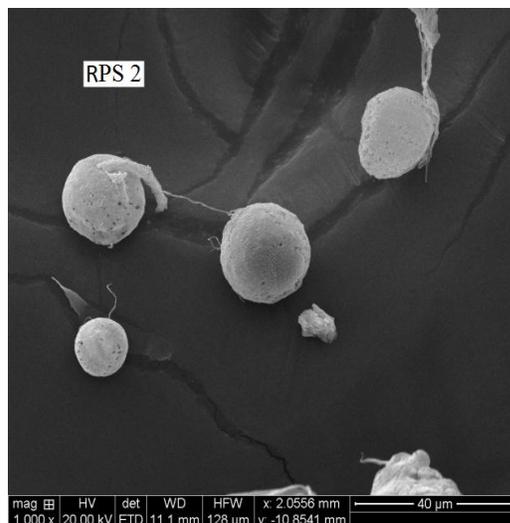
The formulations with HPMC K15 M and EC show angle of repose value in the range of $21^{\circ}.17'$ to $29^{\circ}.23'$ i.e., less than 30° , which shows good flow properties of the formed microparticles

Surface morphology of Rabeprazole Sodium microspheres (SEM)

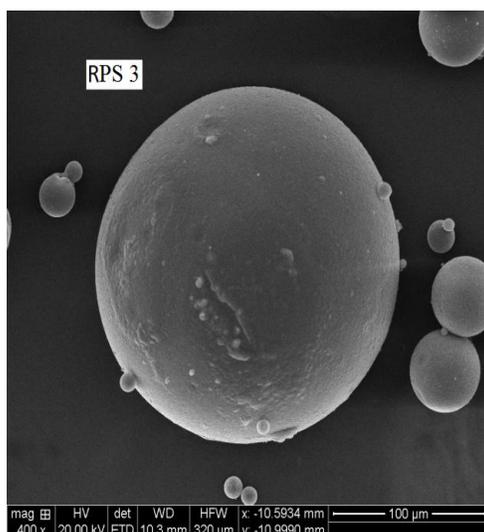
SEM photographs of the various formulations were shown in the Figure 1. Surface smoothness of the RPS floating microspheres was increased by increasing the polymer conc., which was confirmed by SEM. At lower polymer conc. (1:1) rough and wrinkled surface of RPS floating microspheres was obtained in Figure1 and at higher polymer concentration (1:3) the RPS floating microspheres with smooth surface was obtained Figure1. Microspheres with HPMC K15M contain smooth surface and smaller in size compare to the microspheres with ethyl cellulose.



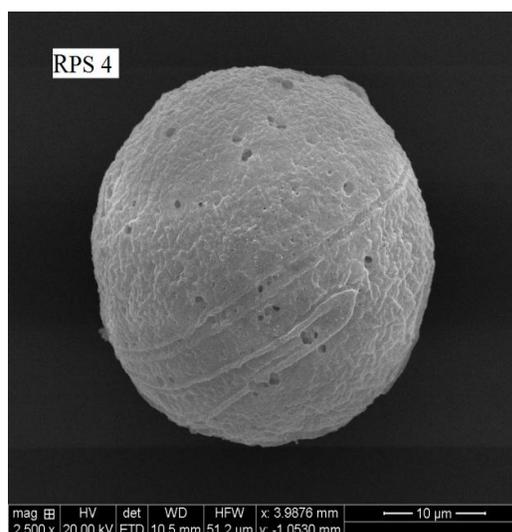
(a)



(b)



(c)



(d)

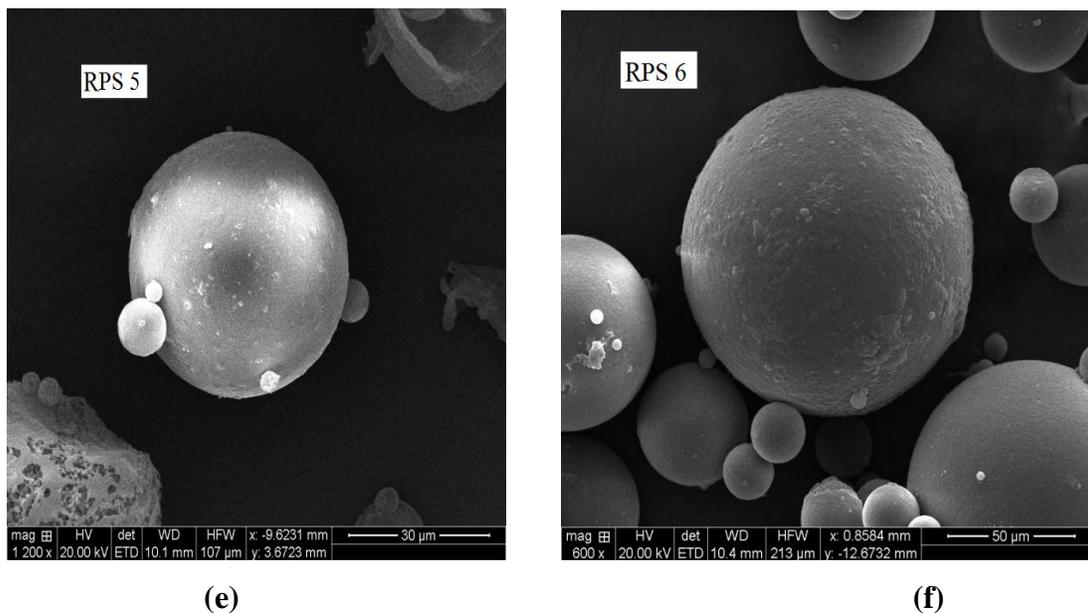


Figure1: SEM photographs of RPS floating microspheres

RPS1, RPS2, RPS3, RPS4, RPS5 and RPS6 refers to RPS floating microspheres prepared by using HPMC K15 M and EC with drug: polymer ratio 1:1, 1:2, 1:3, 1:4, 1:5 and 1:6. HPMC K15M 1,2,3(a, b, c) & ethyl cellulose 4,5,6(d, e, f).

Determination of Average particle size

As the RPS to polymer ratio was increased, the mean particle size of RPS floating microspheres was also increased. The significant increase may be because of the increase in the viscosity of the droplets (may be due to the increase in concentration of polymer solution). RPS floating microspheres with HPMC K15M having a size range less when compared to that of ethyl cellulose which was shown in table 2.

Buoyancy percentage

The microspheres floated for prolonged time over the surface of the dissolution medium without any apparent gelation. As the polymer concentration increases the buoyancy time increases. The results obtain are given in table 2.

Percentage drug entrapment efficiency

Entrapment efficiency increases with increase in the polymer concentration. From the results it can be inferred that there is a proper distribution of RPS in the microspheres and the deviation is within the acceptable limits. A maximum of 89.60% drug entrapment efficiency was obtained in the RPS floating microspheres which were prepared by using HPMC K15 M. A maximum of 83.22% drug entrapment efficiency was obtained in the RPS floating microspheres which were prepared by using Ethyl cellulose. It was further observed that the drug entrapment was

proportional to the RPS: polymer ratio and size of the RPS floating microspheres. By increasing the polymer concentration, the encapsulation efficiency was increased. Results shown in table 2.

Table 2: Average size, Buoyancy percentage and Entrapment efficiency of Rabeprazole Sodium floating microspheres

Formulation	Average size(μm)	% Buoyancy	Entrapment efficiency
RPS1	73.32	78.40 \pm 1.02	70.80 \pm 1.10
RPS2	74.41	83.45 \pm 1.8	78.88 \pm 0.98
RPS3	78.52	91.15 \pm 1.01	89.60 \pm 1.12
RPS4	86.26	67.30 \pm 1.01	53.74 \pm 1.02
RPS5	90.83	74.32 \pm 0.69	66.12 \pm 1.11
RPS6	93.57	79.30 \pm 0.60	83.22 \pm 2.25

***In vitro* dissolution studies**

The *in vitro* performance of RPS floating microspheres showed prolonged and controlled release of RPS. The results of the *in vitro* dissolution studies shows controlled and predictable manner as the polymer concentration increases the drug release from the floating microsphere decreases. The formulations with HPMC K15M i.e.RPS1 95.25% to RPS3 78.35% are shown in Table 3 and Figure 2. The formulations with Ethyl cellulose i.e.RPS4 86.48% to RPS6 64.35% are shown in Table 3 and Figure 3.

Table 3 :*In vitro* release data of Rabeprazole Sodium floating microspheres with HPMC K15M and ethyl cellulose

Time (h)	% Cum. drug release(\pm SD)					
	RPS1	RPS2	RPS3	RPS4	RPS5	RPS6
0	0	0	0	0	0	0
1	17.38 \pm 0.32	13.47 \pm 0.10	12.35 \pm 0.13	21.44 \pm 0.60	10.52 \pm 0.64	7.64 \pm 0.50
2	25.47 \pm 0.22	17.52 \pm 0.16	18.37 \pm 0.20	28.60 \pm 0.49	15.63 \pm 0.57	13.38 \pm 0.54
3	32.49 \pm 0.12	28.31 \pm 0.11	24.31 \pm 0.10	33.50 \pm 0.69	21.50 \pm 0.60	17.78 \pm 0.60
4	39.43 \pm 0.16	32.36 \pm 0.29	30.31 \pm 0.04	38.69 \pm 0.68	27.37 \pm 0.62	23.57 \pm 0.68
5	46.55 \pm 0.13	41.81 \pm 0.23	36.34 \pm 0.26	45.60 \pm 0.67	32.56 \pm 0.59	28.45 \pm 0.60
6	55.32 \pm 0.16	51.27 \pm 0.24	42.33 \pm 0.41	53.68 \pm 0.60	38.40 \pm 0.56	34.01 \pm 0.58
7	64.92 \pm 0.33	56.68 \pm 0.22	48.37 \pm 0.16	60.46 \pm 0.67	43.68 \pm 0.60	38.65 \pm 0.60
8	76.13 \pm 0.26	70.1 \pm 0.12	54.36 \pm 0.07	67.54 \pm 0.58	49.43 \pm 0.56	44.44 \pm 0.69
9	87.77 \pm 0.14	75.62 \pm 0.21	60.37 \pm 0.16	75.63 \pm 0.68	54.62 \pm 0.64	49.52 \pm 0.73
10	92.26 \pm 0.24	83.74 \pm 0.27	66.39 \pm 0.16	80.43 \pm 0.56	60.49 \pm 0.52	54.54 \pm 0.66
11	93.07 \pm 0.42	85.95 \pm 0.18	72.39 \pm 0.36	84.45 \pm 0.59	66.33 \pm 0.60	59.56 \pm 0.56
12	95.25 \pm 0.37	88.98 \pm 0.26	78.35 \pm 0.43	86.48 \pm 0.53	71.55 \pm 0.56	64.35 \pm 0.66

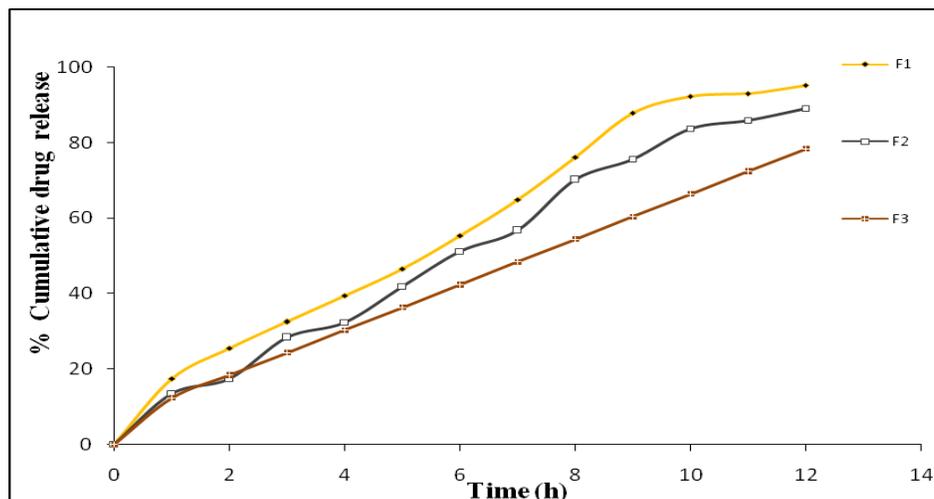


Figure2: Comparative *in vitro* release profile of RPS floating microspheres with HPMC K15M

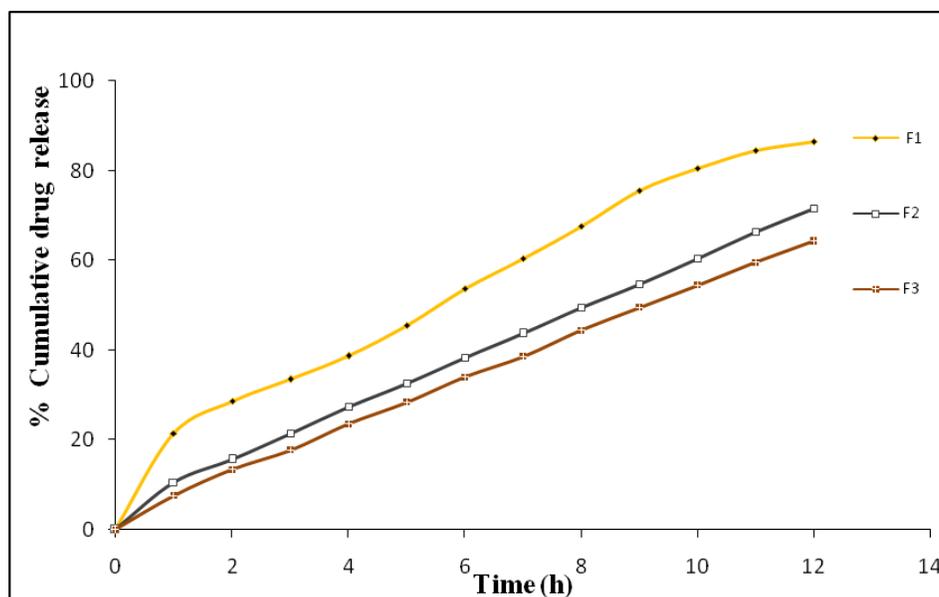


Figure 3: Comparative *in vitro* release profile of RPS floating microspheres with Ethyl Cellulose

Kinetics of drug release

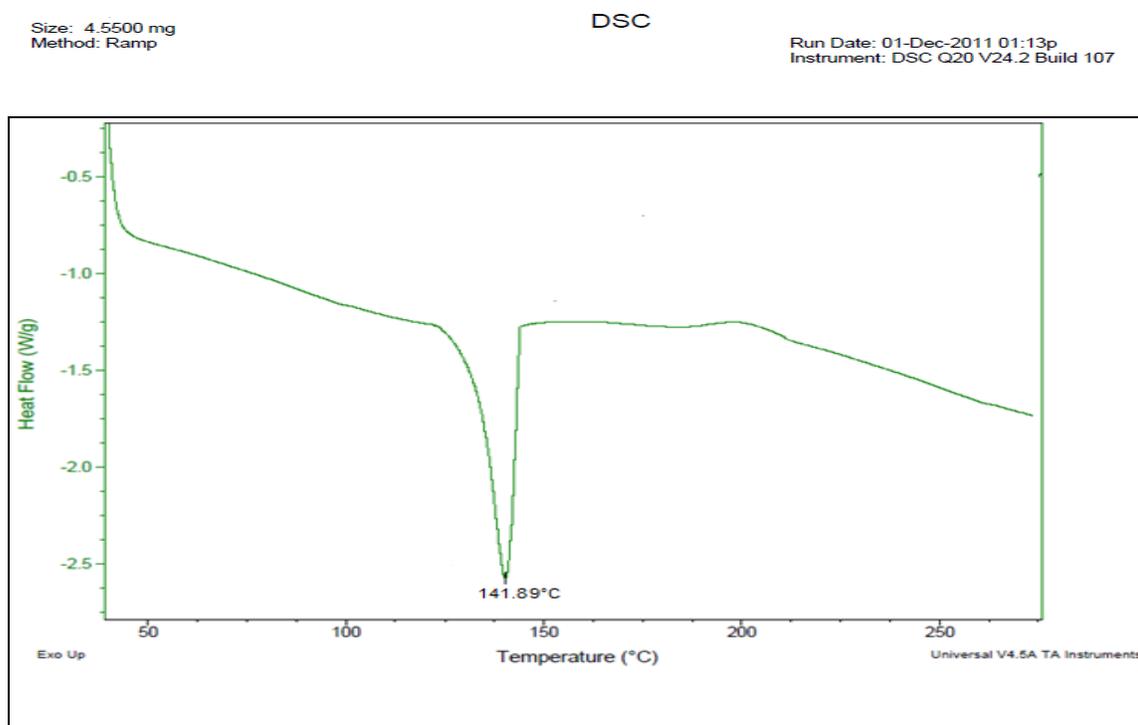
The slopes and the regression co-efficient of determinations (r^2) were listed in Table 4. The co-efficient of determination indicated that the release data was best fitted with zero order kinetics. Higuchi equation explains the diffusion controlled release mechanism. The diffusion exponent 'n' values of Korsmeyer-Peppas model was found to be in the range of 0.5 to 1 for the RPS floating microspheres prepared with HPMC K15 M and EC indicating Non-Fickian of drug through RPS floating microspheres.

Table 4- Pharmacokinetic release of Rabeprazole Sodium floating microspheres with HPMC K15M and EC

Formulation	Zero order	First order	Higuchi Matrix	Peppas plot	
				r ² value	'n' value
RPS1	0.9795	0.9244	0.9473	0.9821	0.7420
RPS2	0.9884	0.9532	0.9376	0.9819	0.8361
RPS3	0.9959	0.9671	0.9764	0.9928	0.7628
RPS4	0.9747	0.9704	0.9721	0.9751	0.6046
RPS5	0.9971	0.9773	0.9454	0.9936	0.7942
RPS6	0.9988	0.9854	0.9378	0.9979	0.8682

DSC Thermograms

In order to confirm the physical state of RPS in the microspheres, DSC of the RPS, RPS loaded floating microspheres with individual polymers were carried out and shown in Fig 4 to 6. The DSC trace of RPS showed a sharp endothermic peak at 141.89°C, its melting point. RPS floating microspheres with HPMC K15M and EC showed the same thermal behavior 147.18°C and 147.32°C respectively indicating that there was no interaction between the RPS and the polymer in the solid state. The melting point range of RPS is between 140-142°C, thus indicating there is no change of RPS in pure state. The absence of endothermic peak of the RPS at 141°C in the DSC of the RPS floating microspheres suggests that the RPS existed in an amorphous or disordered crystalline phase as a molecular dispersion in polymeric matrix.

**Figure 4: DSC of pure drug rabeprazole sodium**

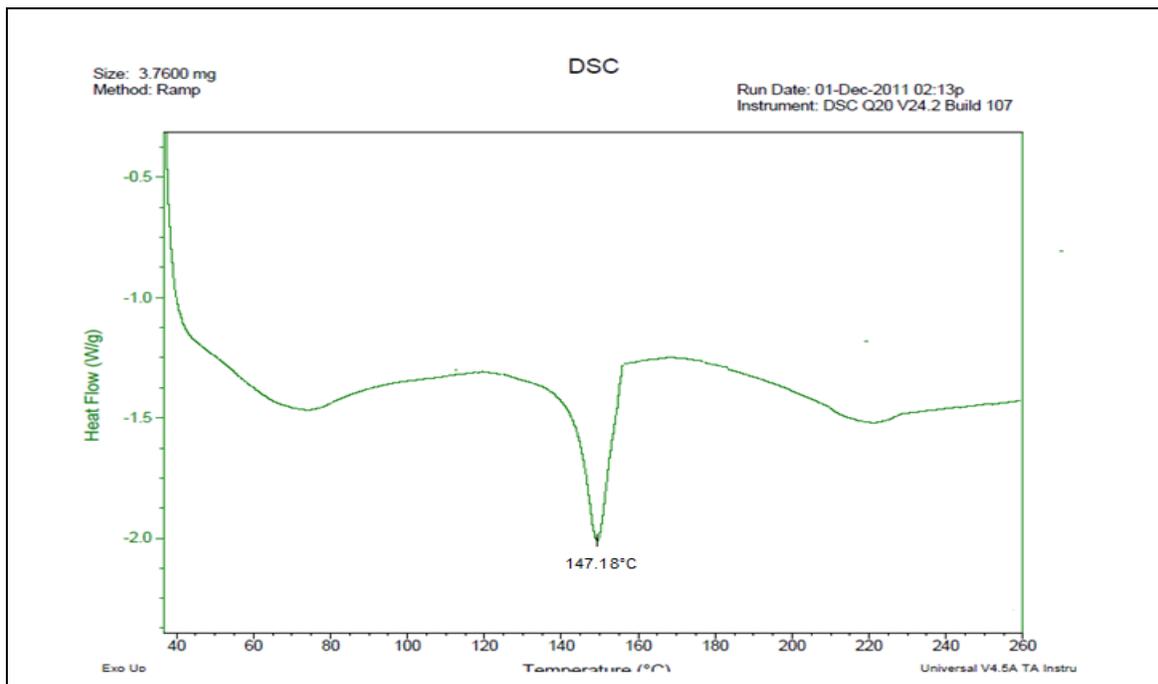


Figure 5: DSC of Formulation with HPMC K15 M

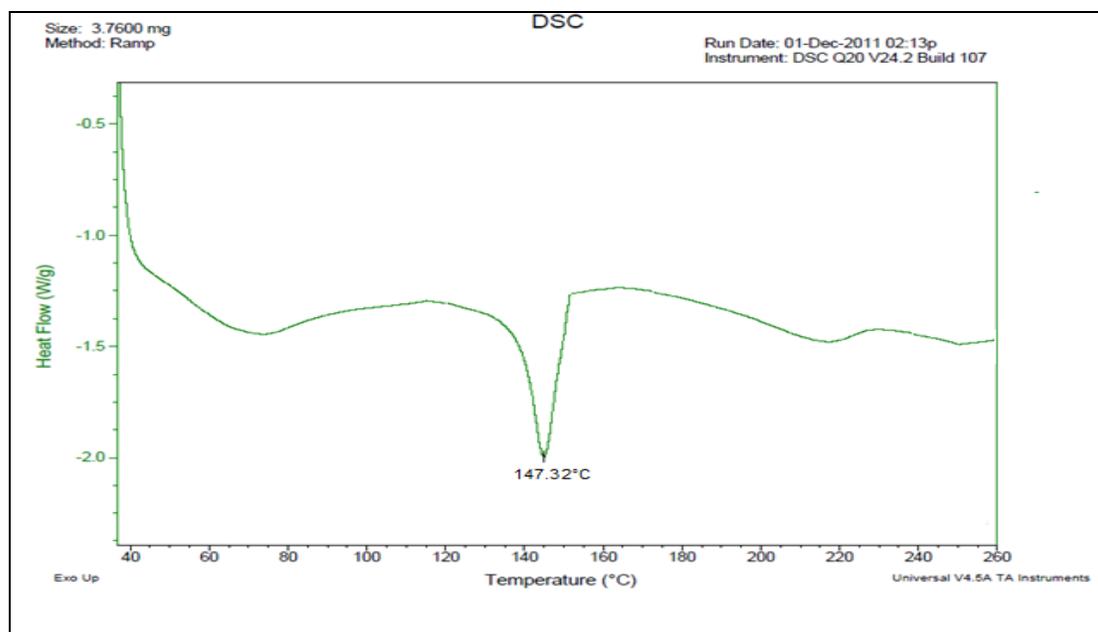


Figure 6: DSC of formulation with ethyl cellulose

X-Ray Diffraction Thermograms

In order to confirm the physical state of the RPS in the microspheres, powder X-ray diffraction studies of the RPS, RPS microspheres with individual polymers were carried out. X-ray diffractograms were shown in Figure 7 to 9 and showed that the RPS is still present in its lattice structure where it is completely amorphous inside the RPS microspheres. This may be due to the conditions used to prepare the RPS microspheres lead to cause complete drug amorphization.

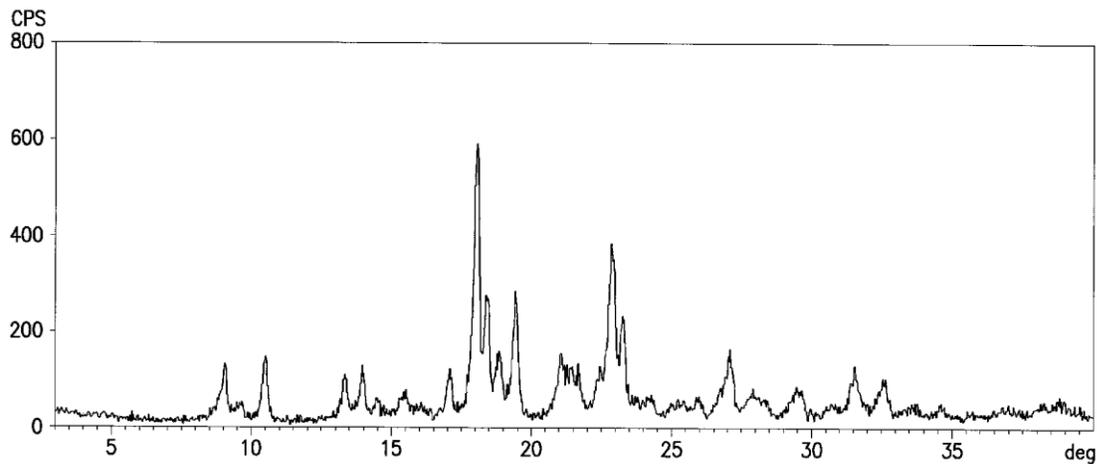


Figure 7: XRD thermogram of RPS

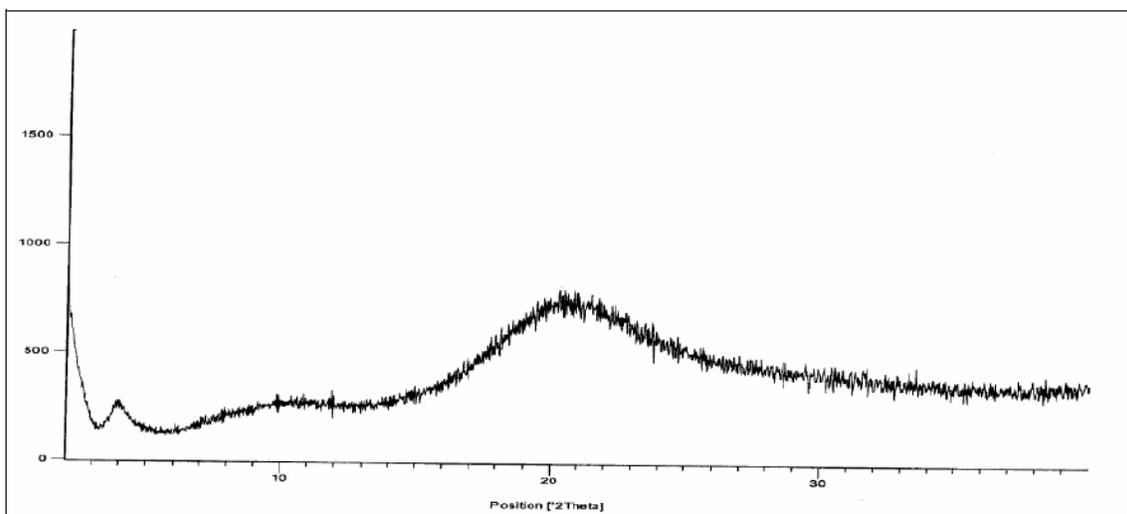


Figure 8: XRD thermogram of RPS formulation with HPMC K15 M

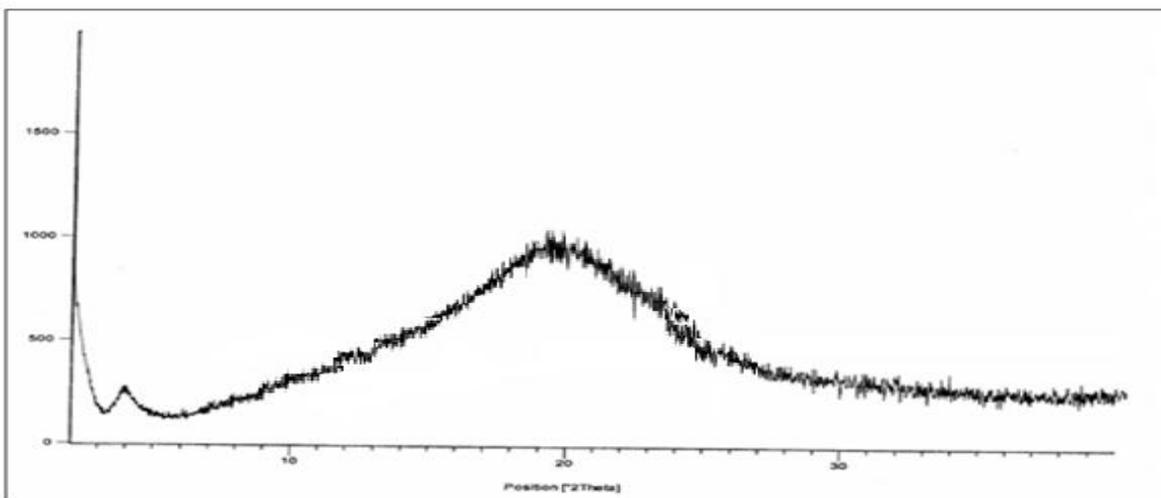


Figure 9: XRD of formulation of RPS with Ethyl cellulose

CONCLUSION

The concept of formulating floating microspheres containing RPS offers a suitable, practical approach to achieve a prolonged therapeutic effect by continuously releasing the medication over extended period of time. floating microspheres of RPS were prepared successfully by emulsion solvent evaporation method using the different concentration of individual polymers like HPMC K15 M, and EC by means of prolonging its gastric retention thus improving the oral bioavailability of the drug. It would be faster and more economical to alter beneficially the properties of the existing drugs than developing new drug entities hence this formulation will be boon to novel drug dosage forms.

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